

The Antimicrobial Effect of The Artimisa Herba and Eordium Glycobhyllum Against A Few of Pathogenic Bacteria

¹Mokhtar Belgacem Halbuda, ²Asma Mohmad Indbha, ^{3*}Mohamed A. Dow

^{1,2}Department of Biology, Faculty of Science and arts (Bader), Al- Gabal Al-Gharby University

^{3*}Department of Microbiology, Faculty of Medicine, Sirte University

Abstract

Artimisa herba and *Erodium antimicrobial* activity were used against *Serratia* spp., *Bacillus cereus*, *E.coli*, *Micrococcus roseus*, *Staphylococcus aureus*, *Streptomyces* spp. and *Micrococcus luteus*. The method of extraction applied in this paper was the Soxhlet- hot extraction using petroleum ether and alcoholic solvents. The bacterial inhibition by the extract was as follows:

Serratia spp was found moderately sensitive to *Artimisa herba* with failure to produce pigment. *B. cereus* was found moderately so to *Artimisa herba* lal, while it showed low response to *Artimisa herba*. *Escherchia coli* showed moderate inhibition to *Artimisa herba*, *Micrococcus roseus* responded well to and *erodium* moderately to *erodium*. *Staphylococcus aureus* well to moderately to *Artimisa herba*. *Streptomyces* spp. responded well only to be based on the results obtained in this study, it can be concluded that using 50% acetone extract of, *erodium glycobhyllum* has high antibacterial activity against pathogenic *Staphylococcus aureus*. Hot Soxhlet extraction method is recommended as well as 96 well trays microtitre plate method. The MIC of tested *Erodium glycobhyllum* against *Staphylococcus aureus* was 25 and 50 mg/ml, respectively. *Peganum* showed moderate response to *Erodium* *Micrococcus luteus* showed height moderate response to *Erodium*, However the exception of *Erodium* in high concentration, none of the tested plants showed any signs of toxicity in rats tested. The former produced liver fatty changes and haemorrhages in the kidneys. Toxicity experiments of *Erodium glycobhyllum*, and *Artimisa herba* using experimental animals (rats) confirm the safety of such plants for therapy of various diseases. An antibacterial evolution for the total extract of tested medicinal plants could also reveal a more potent activity toward tested pathogenic Gram positive and Gram negative bacteria. However, the plants differed significantly in the activity towards tested pathogenic bacteria. The differences could be attributed to their structural nature.

Keywords: *Artimisa herba*, *Eoridium*, Pathgenic bacteria.

1. Introduction

Medical plants are used by 80% of the world population as the only available plants medicines especially in developing countries. In most countries, the percentage of people dependent on medicinal plants for health care is estimated at over 90%. These plants and derived products an important role in the primary health care of Sudan.

in Libya, there are about 1, 825 vascular plant species, of which 134 are endemic. About 450 species are reported to be of medicinal value in the country[1]. Some important families are *Apiaceae*, *Asteraceae*, *Laminaceae*, *Poaceae*, *Fabaceae*, *Brasicaceae* and *Abiaceae*. Medicinal plants are distributed throughout the country, especially in Jabal Al-garbe, Ghedames, Gharian, Awbari and Tarhona district. Many of these plants are associated with a long history of traditional use. More than a hundred species are massively used by Bedouins or local people in folk medicine as hot or cold drinks or chewed fresh or dry. These are also used externally to cure dermal diseases, viral diseases and bacterial infections, insect or animal bites or burns and sometimes for the treatment of hair problems. These medicinal plants are very well documented in different flora. Many species of medicinal plants, such as *Asperula arvensis* L., *Cupressus sempervirens* L., *Juniperus phoenicea* L., *Pinus halepensis* Mill., *Quercus coccifera* L., *Tribulus longipetalus* Viv., *Veronica cymbalaria* Bodarod and *Vahlia dichotoma* (Murray) Kuntze have become threatened by extinction [2,3]. Owing to over-harvesting and diversion of forest land containing medicinal plants to agriculture .

The best known medicinal plant of Libya is *Silphium cyrenaicum* It is existed during Greek and Roman times. The plant was used for the treatment of many illnesses and was so important to the economy of the country that it used to be sold for its weight with silver or gold and its picture was depicted on currency coins.

There are many kinds of flower vegetables that have medicinal properties. These are flowers of traditional vegetable plants that have flowers and have also been used for cooking since ancient time. It was believed that consumption of these flowers vegetable can cure illness and infectious diseases. They also help people who suffer from diarrhea, which indicates the antimicrobial activity of these vegetables [4,5,6]. In addition, the

countries in central Africa, West Africa and Indonesia used "*khilck*" flower (*Senna siamea*) to treat patients who had stomachache [7].

The flower vegetables are composed of various chemical compounds that can be grouped as flavonoid, anthraquinone and glucoside-flavonoid is expected to be the main component that plays a major role in microbial inhibition [8]. Even essential oils were found to have inhibitory effect on pathogenic bacteria and some were found to have antimalarial and antimicrobial property [9,10].

Herbs and spices along the Mediterranean sea have been shown to possess antimicrobial actions and could serve as a source for antimicrobial agents against food spoilage and microbial pathogens [11,12,13,14,15,16].

(1) *Artimisa herba*

(2) *Eroduim glauchophllum*

1- Artimisa herba:

Artemisia genus asso (*compositae*) include 15 perennial aromatic herbs and shrubs that grow wild in dry or semi-dry habitats in the provine of Alberta , Canada and in Libya [17]. Asso. (syn : *A. incutta* Del., commonly known as the desert worm wood (*Arabic name : Sheeh*) , is a dwarf shrub or semishrub) growing widely in Jordan and in the Middle East . The plant is a perennial, strongly aromatic herb, with many basal , erect and leafy stems covered by woolly hairs . The flower heads are about 0.5 cm in diameter, not showy, and made only of tubular florets. The plant is mainly used as anthelmintic and antispasmodic and for other common uses in folk medicine such as relief of coughing , intestinal disturbances , colds and muscle relaxant by the local population [18,19].

Previous works have suggested that several suggested oils showed important antimicrobial activity against bacteria, yeas, dermatophyte and *Aspergillus strains* [20,21,22] and have therapeutic potential , mainly in diseases involving mucosal, cutaneous and respiratory tract infections. The major constituents of many of these oils are phenolic compounds (terpenoids and phenyl propanoids) like thymol, carvacrol or eugenol, of which antimicrobial and antioxidant activities are well documented [20, 22].

Nevertheless, aromatic plants producing non – phenolic essential oils, like some *Artemisia*

species, are also used as spices and in folk remedies as antiseptics. Powdered leaves of *A. absinthium*, *A. biennis*, *A. frigida* and *A. ludoviciana* have been applied externally in salves and washes by North American native people for treating sores and wounds and, internally to treat chest infections [23]. In addition, *A. herba-alba* possessed antimicrobial activity against some pathogenic bacteria and yeasts using the broth micro-dilution method [17].

Recently, several species of Artemisia essential oils had inhibitory effect on the growth of bacteria (*Escherichia coli*, *Staphylococcus aureus* and *Staphylococcus epidermidis*), Yeasts (*Candida albicans*, *Cryptococcus neoformans*), dermatophytes (*Trichophyton rubrum*, *Microsporum canis*, and *microsporum gypseum*) *Fonsecaea pedrosoi* and *Aspergillus niger* [24].

Additionally to the antimicrobial activity of *Artemisia herba-alba* Asso, [25]. In Jordan reported that the essential oil of the plant promising candidates as natural herbal constituents of antimicrobial drug combinations. In Morocco, [26]. Concluded that *Artemisia herba-alba* Asso. aqueous extract possesses antihypertensive activity in spontaneously hypertensive rats and that the underlying mechanism appears to involve, at least in part, an increase in urine and electrolyte output. However, in Tunisia, it was also reported [27].

Artemisia oils of some species had inhibitory effects on the growth of bacteria (*Escherichia coli*, *Staphylococcus aureus*, and *Staphylococcus epidermidis*), Yeasts (*Candida albicans*, *Cryptococcus neoformans*), dermatophytes (*Trichophyton rubrum*, *Microsporum canis* and *microsporum gypseum*), *Fonsecaea pedrosoi* and *Aspergillus niger*. *Artemisia biennis* oil was the most active against dermatophytes, *Cryptococcus neoformans*, *Fonsecaea pedrosoi* and *Aspergillus niger*, and *Artemisia absinthium* oil the most active against *Staphylococcus strains* [24].

2- *Erodium glaucophyllum* (Ebrat Elrahib) :

The family Geraniaceae encompasses many promising plants from the medicinal plant of view. Some of the family members are specified in the Chinese pharmacopoeia and formulated in the Chinese herbal medicine such as Lao-guan-cao formula. The drug consists of *Erodium stephanianum*, *Geranium nepolense* and *Geranium sibiricum* and is

used to promote circulation in acute and chronic rheumatologic disorders and as detoxicant for enteritis and basillary dysentery [28]. From the chemical point of view , caffeine , tyramine , glutamic acid , choline , gallic acid , saponins , flavenoids and sugar were reported as common active constituents of *Erodium cicutarium* as well as of the family Geraniaceae [29]. Results of the antimicrobial activity against *Escherichia coli*, *Staphylococcus aureus* and *Candida albicans* proved comparable activity in comparison to those of ampicillin, gentamycin and mycostatin . The results of the MIC values after 48h incubation at 37C° were determined by two-fold serial dilution assay [30]. The natural susceptibility to 71 antibiotics of 104 *S. Serratia ficaria* , *Serratia rubidaea* , *Serratia fonticola* , *Serratia odorifera* , and *Serratia plymuthica* was examined by Stock and his colleagues[31].

In this study it was found valuable to screen some of the wild medicinal plants such as *Artimisa Helichrysum stoechas* and *Erodium glycyphyllum* from Libya against some Gram-positive and Gram-negative pathogenic bacteria such as: *Staphylococcus aureus* ,Micrococcus species, *Bacillus cereus* , *Serratia species* , *Escherichia coli* , *Salmonella species* and *Pseudomonas species*.

2. Materials and Methods

1- Plant Materials:

a. *Artimisa herba*.

Artimisia is a large, diverse genus of plants with between 200 to 400 species belonging to the daisy family Asteraceae. It comprises hardy herbs and shrubs known for their volatile oils. They grow in temperate climates of the Northern Hemisphere, usually in dry or semi-dry habitats. The fern-like leaves of many species are covered with white hairs. Common names used for several species include wormwood, mugwort, sagebrush and segewort, while a few species have unique names, notably Tarrago (*Artemisia dracunculus*) and Southernwood (*A. abortanum*). Occasionally some of the species are called sages, causing confusion with the saliva sages in the family Lamiaceae. Artemisia species are used as food plants by the larvae of a number of Lepidoptera species.

Artemisia genus as so (composite) include 15 perennial aromatic herbs and shrubs that

grow wild in dry or semi-dry habitats in the province of Alberta, Canada and in Libya [17]. About 400 species, mostly distributed in North temperate parts of the world (common on arid soils and halophytic habitats), South America and South Africa; 5 species are reported from Libya.

Artemisia herba-alba is one growing species in Libya. It is aromatic low growing, much branched from the base, up to 30 cm high shrubby perennial; stems usually patent, at first densely cane scented later becoming glabrescent below.

The plant is distributed in Wadi louti, Gharian, mountains, sand stone and limestone, common in Yefren, Wadi Ain Romia, Alghazaya towards Wazzin. Additionally to Libya, *Artemisia herba-alba* is distributed in West Southern France, Spain, North Africa, Syria extending into West and Central Iran.

2- Preparation plant extracts:

a. Collection of plants.

a-1. Collection of plants :

The plants were collected during the spring season of 2012 from AL- Gabal AL- Gharbie mountain Libya. The plants were classified and authenticated according to Jafri and El-ghadi (1978). In Department of pharmacognosy, Faculty of pharmacy, Tripoli University, Tripoli, Libya [32].

a-2. Extraction of tested plants

The methods for preparation extracts mainly organic solvents according to (Wagner and Bladt, 1995).

a-3 Activity of tested plant extracts :

The tests were performed by the Muller Hinton agar well diffusion method [33]. *Streptococcus* species, *Serratia* species and Gram-positive strains; *Staphylococcus aureus*, *Micrococcus* species, *Bacillus cereus*. The tested strains of pathogenic bacteria used in this investigation are human isolates obtained from Research Center of Biotechnology, Tripoli, Libya [34]. To dissolve and dilute the extract, 50% acetone at proper concentration was used. Control plates containing only 50% acetone. Tests were performed in duplicate.

a-4 Agar diffusion cup cut method ; Eight serial dilutions generating decreasing concentrations of 100, 50, 25, 12.5, 6.25, 3.125, 1.56 and 0.78 mg/ml for the acetone extract

of Artemisia herba alba and Erodium glaucophyllum were first performed..

3. Results

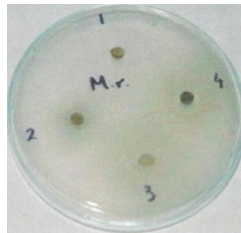
I- Screening Antibacterial Activity of some Medicinal Plants against certain Pathogenic Bacteria :

In order to test optimum condition of tested plant extraction for antibacterial activity, different solvents with increasing polarity such as water and ethanol were used. Water was used in aqueous extraction as well as ethanol. Crude extracts yield of shoot system are shown in Table (1). As shown in the same table the crude ethanolic extraction of Artimisa herba was higher than crude *Erodium* water extracts. However, water extract appears higher solvent for Artimisa herba. This was primarily studied against certain pathogenic bacteria such as *Serratia sp.*, *B. cereus*, *E. coli*, *Micrococcus aureus*, *Streptomyces sp.* and *Micrococcus luteus*. The data in table shows that failure of pigment formation of red colour in growth surrounding the clear zone of *Serratia sp.* under the effect of water extract. The preliminary results of antimicrobial activity test varied from low to high inhibition of growth against tested pathogenic bacteria by two solvents used (water and ethanol) as shown in Table (1). The aqueous extraction generally gave less antibacterial activities than ethanolic extraction. The ethanolic extraction of the shoot systems from tested medicinal plants produced high inhibition zone against certain pathogenic bacteria. Aqueous extract of *Artimisa herba* gave moderate inhibition against tested bacterial species. So it is considered that the aqueous extract of tested plants displayed no antibacterial activity against tested pathogenic bacteria.

Table 1. Growth of different bacterial spp. In the presence of alcoholic and water extracts of some medicinal plants

Tested plants	<i>Erodium Glucophyllum</i>		Artimisa herba	
	Alcohol	Water	Alcohol	Water
<i>Serratia sp.</i>	-	-	+*	++*
<i>B. cereus</i>	++	-	+	++
<i>E. coli</i>	++	-	++	++
<i>Micrococcus raseus</i>	+	-	++	+
<i>Staphylococcus aureus</i>	-	-	-	++
<i>Streptomyces sp.</i>	+++	-	-	++
<i>Micrococcus Luteus</i>	++	-	+++	++

: Failure in pigment formation in the growth surrounding the clear zone. : Negative inhibition, + : Low inhibition, ++ : Moderate inhibition, +++ : High inhibition



1- Ethanolic extract of Erodium



2- Aqueous extract of Erodium



3- Ethanolic extract of *Artemisa herba*



4- Aqueous extract of *Artemisa herba*

M.r. : *Micrococcus roseus*

Staph. : *Staphylococcus aureus*

M.L. : *Micrococcus loteus*

E.C. : *Escherichia coli*

B.C. : *Bacillus cereus*

Strepto. *Streptomyces sp.*

Antibacterial activity of some selected medicinal plants :

In this experiment we used two plants namely; *Artemisa herba*, and, *Erodium glycyphylum* Using 50% acetone as solvent system, according to **Khond et al. (2009)**. The highly pathogenic bacterial strains tested were *Escherichia coli*; *Salmonella sp.*, *Pseudomonas sp.* and *Staphylococcus aureus*. The obtained results revealed that 50% acetone was suitable solvent which has an antibacterial activity towards Gram +ve *Staphylococcus aureus* with a wide zone of inhibition at a concentration of 100 mg *Erodium glycyphylum*. Table(2). Our results showed that the 50% acetone was the only effective solvent, therefore further investigations were carried out. Other tested strains was tried which had narrow zone of inhibition at concentration 100 mg of either *Erodium glycyphylum*.

Table 2. Antibacterial activity of *Erodium glycopyllum* extracts using 50% acetone.

<i>Bacte Bacterialstrains</i>	Solvent systems			
	Con	Contralrol	55	55% Acetone
<i>Esch erichia coli</i>	-ve-	-ve	-ve	-ve
<i>Salmo Salmonella sp.</i>	-ve	-ve	-ve	-ve
<i>Sta Stahylococcus aureus</i>	-ve	-ve	+ve	+ve
<i>Pseu Pseudomonas</i>	-ve	-ve	-ve	-ve

-ve = no effect

+ve = effect

III- Determination of minimum inhibition concentration (MIC) of 50% acetone extract:

1- Broth microdilution method :

Using the 96 well trys (Fig. I), 100 ul of 50% acetone extract of *Erodium glycopyllum* diluted two fold in decreasing concentration and 5 ul of bacterial inoculums of *Staphylococcus aureus* was distributed in the wells.

Table (1) illustrate that 25 mg/ml of acetone extract is the MIC for *Staphylococcus aureus*. However, Table (2) shows that 50 mg/ml of acetone extract is the MIC for *Staphylococcus aureus* while 100 mg/ml of *Erodium glycopyllum* extract was bactericidal for the same species.

Table 3. Determination of MIC of acetone extract of *Erodium glycopyllum* by Broth micro dilution method.

Bacterial strains	Visible turbidity				MIC
	100 mg/ml	50 mg/ml	25 mg/ml	12.5 mg/ml	
Staphylococcus aureus	clear	clear	turbid	turbid	50 mg/ml

Effect of Tested Plant extract on Experimental Animals:

Artimisa herba

The treated rats were closely observed for 24 hours for any behavioral changes or abnormal clinical signs. Visual observation revealed that no any abnormal behavior seen, no mortality on all doses used. Postmortem was done on the second day of the experiment, in all groups no pathological changes seen in any of the vital organs(liver and kidney).

Table 4. Mortalities and Post-mortem findings in rats treated with *Erodium glaucophyllum*

Gr	OUP	Concentration mg/kg	No. of Rats	% Mortality After 24 hrs	Survival Rate	Mortality (%)	Changes in Tissue	
							Liver	Kidney
1	250 (mg/kg)	6	0	6	0	-ve	-ve	
2	500 (mg/kg)	6	0	6	0	-ve	-ve	
3	1000 (mg/kg)	6	0	6	0	++	-ve	
4	2000 (mg/kg)	6	4	2	66.66	+++	++	
5	Control	6	0	0	0	-ve	-ve	

(-ve): No changes, (++): Fatty changes, **(+++): Extensive Fatty changes.**

4. Discussion

Medicinal plants, since immemorial times, have been used in virtually all cultures as a source of medicine. The widespread use of herbal remedies and healthcare preparations obtained from commonly used traditional herbs and medicinal plants have been traced to the occurrence of natural products with medicinal properties.

The discovery of new drugs from natural compounds has been of significant importance in the past and obviously will continue to be just as important if not more in the future.

Plants are the oldest source of pharmacologically active compounds, and have provided human kind with many medically useful compounds for centuries. Some 7000 medicinal compounds used in the Western pharmacopoeia are derived from plants. Hence, screenings of antimicrobial plants for new agents poses an enormous challenge and are important especially with the emergence of drug resistant diseases.

Additionally to the *Artemisia* ethanolic extract showed antibacterial activity

against *B. cerius*, *Micrococcus roseus*, *E. coli*, *Streptomyces sp.* and *Micrococcus loteus*. This results supported the findings of Dababneh [36]. He reported that *Artemisia-herba-alba* had marked significant sensitivity towards pathogenic *Staphylococcus aureus*, *Pseudomonas aeruginosa*, *E. coli* and *Candida albicans* using well-plate diffusion method. Recently, Hudaib and Aburjai [25] reported that the oil of Jordanian species of *Artemisia* promise candidates as natural herbal constituents of antimicrobial drug combinations. In addition, others stated that *Artemisia sp.* extract is anti-diabetic, antitumor, antifungal against *Candida albicans* and bactericidal[36].

Results of antibacterial activity of *Erodium glycyphillum* exhibit positive effect against *Staphylococcus aureus* at 100 mg. This obtained results has confirmed earlier by Gohar et al. [30]. They reported that extract of *Erodium* has antibacterial activity against *Staphylococcus aureus*, *Escherichia coli* and *Candida albicans*. Despite of the the absence of tannins in *Erodium* reported by others [30,37] proved that *Erodium glycyphillum* contains such constituent. Moreover, the use of some plants of the family *Geraniaceae* for treatment of enteritis and basillary dysentery [28] suggested the presence of antibacterial principles. Fecka et al. [38] reported that some species of the *Erodium genera* contains six natural polyphenolic compounds, brevifolin carboxylic acid, brevifolin, ellagic acid, methyl gallate, gallic acid and protocatechuic acid in methanol extract of the whole plant of *Erodium cicutarium*. Accordingly, they concluded that *Erodium* has an active component called geraniin. From the biological point of view, geraniin was reported to exhibit inhibitory activity for chitin synthase II, inhibit the level of serum cholesterol, glutamate pyruvate transaminase and inhibits the formation of 5-lipoxygenase which should be a useful principal to find a natural drug for asthma and inflammation[39].

Results from this study indicated that tested extracts possessed variable antimicrobial effects against Gram-positive and Gram-negative bacteria. However, the plants differed significantly in the activity against tested microorganisms. These differences could be attributed to the structural nature of plant constituents and microorganisms as well as concentration used. The optimal effectiveness of medicinal plants may not be due to one active constituent, but to the combined actions of different plant constituents. Moreover, the difference observed in antimicrobial activities of the investigated plant extracts suggest the

susceptibility variations of microorganisms to various chemical components. Earlier results support our findings that the compositions of essential oils depend on the plant species, the chemotypes and the climatic conditions, which lead to variation of their antimicrobial activities.

It could be concluded that the tested plant extracts exhibits a broad spectrum of activity against various microorganisms. Results of the present study should be considered for the possible application of plant extracts of natural bacteriostatic and bactericidal component in various products and as natural preservatives extending the pharmaceutical and deity products shelf life, as we believe is of great importance.

In the case of *Erodium* and its effect on rats, it's clear that lower concentrations did not have any deleterious effects on the rats, a part from slight fatty changes and haemorrhage in both kidneys and livers. However, when higher concentrations were used *viz*: 2000 mg/kg 66.67 % of rats died. Dead animals showed liver fatty changes and haemorrhage. Similar lesions were observed in the kidneys.

As far as *Artemisia herba*, all concentrations didn't show any visible lesions which ensure the safety of using such medicinal plants for therapy of various diseases.

5. References

- [1] Auzi , A . (1999) : Medicinal plants in Libya . First Conferences on Natural Resources in Sert , Libya .
- [2] Faraj , I . ; Missaoui , M . ; Bougrine , H . ; Jebriel , A . and Michri , M . (1988) : Evaluation study of Economic value of forest tree in Libya .
- [3] Al-Idrissi , M . ; Sbeita , A . ; Jebriel , A . ; Zintani , A . ; Shreidi , A . ; Ghawawi , H . and Tazi , M . (1996) : Country Report to the FAO international Technical Conf . on Plant Genetic Resources , Leipzig , Germany .
- [4] Somanapan , A . (1990) : Medicine from herbal plant . 1st ed. Faculty of Pharmacy , Mahidol University , Bangkok , Thailand . *Staphylococcus aureus* infection . N . Engl . J . Med . , 340 : 517 – 523 .
- [5] Vachirasup , T . (1995) : Senna plant in Thailand . 1st ed. Faculty of Pharmacy , Mahidol University , Bangkok , Thailand . Warren E . Lewinson & Ernest Jawatz , chapter 10 . Medical Microbiology & Immunology , 3rd ed. Appleton & Lange , Norwaik , Connecticut , 42 – 52 .
- [6] Boonyaprapatsara , N . (2000) : Thai traditional Herbal Medicine plant . Vol . 4 Prachachon Publ . Bangkok , Thailand .

- [7] Grieve , H . (1981): The herb : New plants [http : www : Botanical . com \ Botanical . \ mgmh \ comidx . Html](http://www.Botanical.com/Botanical/mgmh/comidx.html) .
- [8] Proestos , C . ; Boziaris , I . S . ; Nychas , G . J . E . and Komaitis , M . (2006) : Analysis of flavenoids and phenolic acids in Greek aromatic plants , Investigation of their antioxidant capacity and antimicrobial activity . *Food Chem* . 95 : 664 – 671
- [9] Oussalah , M . ; Caillet , S . ; Saucier , L . and Lacroix , M . (2006) : Antimicrobial effects of selected plant essential oils on the growth of *Pseudomonas putida* strain isolated from meat . *Meat Science* , 73 : 236 – 244
- [10] Van Vuuren , S . F . ; Viljoen , A . M . ; Van zyl , R . L . ; Van Heerden F . R . ; Husnu , K . and Basser , C . (2006) : The antimicrobial , antimalarial and toxicity profiles of helihuinulone , leaf essential oil and extracts of *Helichrysum cymosum* L. South Africa of Botany , 72 : 287 – 290 .
- [11] Conner , D . E . and Beuchat , L . R . (1984) : Effects of essential oils from plants on growth of food spoilage Yeast . *J . of food Science* , 49 : 429 – 434 .
- [12] Zaika , L . L . (1988) : Spices and Herbs : Their antimicrobial activity and its determination . *J . of food safety* , 9 : 97 – 119 .
- [13] Conner , D . E . (1993) : Naturally occurring compounds . In : P . Davidson and A . L . Branen (eds .) . antimicrobials in foods (pp . 441 – 468) New York , N Y : Moreel Dekker .
- [14] Dorman , H . J . and Deans , S . G . (2000) : antimicrobial agents from plants : Antimicrobial activity of plant volatile oils . *J . Appl . Microbiol* . 88 : 308 – 316 .
- [15] Bagamboula , C . F . ; Uyttendaele , M . and Debevere , J . (2003) : Antimicrobial effect of spices and herbs on *Shigella sonnei* and *Shigella flexneri* . *J . of food protection* , 66 : 668 – 673 .
- [16] Mouuia , O . ; Stephane , C . ; linda , S . and Monique , L . (2006) : Inhibitory effects of selected plant essential oils on the growth of 4 pathogenic bacteria : *E . Coli* , O157 : H7 , *Salmonella typhimurium* , *Staphylococcus aureus* and *Listeria monocytogenes* . *Food control* , [www . Elsevier . Com \ Locate \ food count](http://www.Elsevier.Com/Locate/foodcount) .
- [17] Bogdadi ,H. A. A.; Kokoska, L. ; Havlik, J.; kloucek , P. ; Rada , V. and Vorisek, K. (2007) : In vitro antimicrobial activity of some Libyan medicinal plant extracts . *Pharmaceutical Biology* , 45 (5) : 386 – 391 .
- [18] Abu-Irmailehand , B. and Afifi , F. (2003) : Herbal medicine in Jordan with special emphasis on commonly used herbs *J. Ethnopharmacol.* (89) : 193 – 197
- [19] Oran , S . A . and Al-Eisawi , D . M . (1998) : Check-list of medicinal plants in Jordan .*Dirasat Med. Biol. Sci.* , 25 (2) : 84 – 112 .
- [20] Griffin , S . G . ; Wyllie , G . ; Markham , J . L . and Leach , D . N . (1999) : The role of structure and molecular properties of terpenoids in determining their antimicrobial activity . *Flav . fragr . J .* 14 : 322 – 332 .
- [21] Rios , T . ; Recio , M . and Villar , A . (1990) : Antibacterial activity of *Helichrysum stoechas* . *Planta Med.* 56 : 446 .
- [22] Janssen , A . M . ; Scheffer , J . J . C . ; and Svendsen , A . B . (1987) : Antimicrobial activity of essential oils : a 1976 - 1986 literature review . *Aspects of the test methods . Planta Med .* 5 : 395 – 398 .
- [23] Kreshaw , L . (2000) : *Edible and medicinal plants of the Rockies . Lone Pine Edmonton . Canada .*
- [24] Lutz , D . L . ; Alviano , D . S . ; Alviano , C . S . and Kolodziejczyk , P . P . (2008): Screening of chemical composition, antimicrobial and antioxidant activities of *Artemisia* essential oils . *Phytochemistry* 69 : 1732 – 1738 .
- [25] Hudaib , M . M .and Aburjai , T . A . (2006) : composition of the essential oil from *Artemisia herba-alba* grown in Jordan . *J . of Essential oil Res . , (JEOR)* pp. 18 – 23

- [26] Zeggwagh , N. A.; Farid , O. , Michel , J. B. and Eddouks , M. (2008) : Cardio-vascular effect of *Artemisia herba-alba* aqueous extract in spontaneously hypertensive rats . Methods find Exp. Clin. Pharmacol. 30 (5) : 375 – 381 .
- [27] Ben Abid , Z. ; Feki , M. ; Hedhili , A. and Hamdaouil ,M. H. (2007) : J. of Nutrition , Metabolic Diseases and Dietetics 51 (3) : 16 .
- [28] Zhang , Y.Y; Li, S.H. and Tian, Z.(1995) : Morphological and Histological studies of the Chinese drug Lao-guan-cao . Yao xue Bao 3 : 46 – 58 .
- [29] Hussein, F. T.K. (1985) : Medicinal plants in Libya . 1st ed. Arab Encyclopedia House, Beirut, Lebanon. P. 436 .
- [30] Gohar, A. A.; Lahloub, M.F. and Niwa, M. (2003): Antibacterial polyphenol from *Erodium glaucophyllum*. Z. Naturforsch. 55 : 670 – 674 .
- [31] Stock , I. ; Burak, S. ; Jane, K.; Gruger, T. and Wiedemann, B.(2003B) : Natural Antimicrobial Susceptibilities of strains of “unusual” *Serratia species* : *S.ficaria*, *S.fonticola*, *S. odorifera*, *S.plymuthica* and *S. rubideae* . J. Antimicrobial Chemotherapy 51 : 865 – 885 .
- [32] Jafri , S. and El-Gadi , A. (1978) : *Asteraceae* . In flora of Libya , 107. Al-Fateh University Press , Tripoli, Libya .
- [33] Perez, C.; Paul, M. and Bazerque, P. (1990): An antibiotic assay by agar well diffusion method. Acta Biol. Exper. 15:113-115.
- [34] Valgas, C. ; Machado de Sousa, S. ; Smania, FA. E. and Arthur Smania, Jr. A. Jr. (2007) : Screening methods to determine antibacterial activity of natural products. Brazillian Journal of Microbiology 38: 369-380.
- [35] Khond, M. ; Bhosale, J.D.; Arif, T.; Mandal, T.K.; Padhi, M.M. and Dabur, R. (2009) : Screening of some selected medicinal plant extracts for In-vitro Antimicrobial activity. Middle-East J. Scientific Res. 494) : 271-278.
- [36] Dababneh, B.F. (2008) : Antimicrobial activity of selected Jordanian medicinal plant extracts against pathogenic microorganisms. J. Food Agric. And Environm. 6(2) : 134-139.
- [37] Gibbs , R.D. (1974) : Chemotaxonomy of flowering plants . Vol. III.Mc Gill Queen's University Press, London. pp. 1337 – 1338 .
- [38] Fecka, I. ; Kowalczyk, A. and Cisowski, W. (2001) : Phenolic acids and Depsides from some species of the *Erodium* genera. Z. Naturforsch., 56c : 943-950.
- [39] Kumari, Y. ; Okuda, H. ; Okuda, T. and Arichi, S. (1986) : Studies on the activities of tannins and