# Solid Phase Extraction Preconcentration and HPLC Determination of Biogenic Amines Using A Hydrazone Derivative Immobilized on Sol-Gel Matrix

Abdassalam A. Alfergani<sup>\*1</sup>, Ali E. M. Alzarqah<sup>2</sup> and Eshtawe M. Agadid<sup>3</sup>

E-mail: salamtameem@yahoo.com

<sup>1,3</sup>Faculty of Education, Department of Chemistry, Sirte University, Sirte, Libya <sup>2</sup>Faculty of Education, Department of biology, Sirte University, Sirte, Libya

#### Abstract

A sorbent material based on a hydrazone ligand, formaldehyde-2,4–dinitrophenylhydrazone was prepared by immobilizing the ligand into a silica sol-gel matrix. The capability of the sorbent material for the extraction of seven biogenic amines was investigated. Under the optimum condition, the sorbent material exhibited preferential selectivity towards the aliphatic (putrescine, PUT; cadaverine, CAD; spermidine, SPD) and the heterocyclic (histamine, HIS) over the aromatic (tryptamine, TRY; 2-phenylethylamine, PEA, tyramine, TYR) biogenic amines.

Keywords: Hydrazone, biogenic amine. Sol-gel, extraction

## 1. Introduction

Biogenic amines (BAs) are produced in plants and animals by microbial decarboxylation of amino acids resulted from removing  $\alpha$ -carboxyl group to produce the corresponding amine. BAs can have an aliphatic, aromatic or heterocyclic structure, and their study is of interest for their role as biomarkers in toxicological risk studies and as indicators of food quality [1]. They have also toxicological effects but the exact toxicity threshold values for compounds are different, since the toxic dose is strongly dependent on the efficiency of the detoxification mechanisms of each compound. A considerable number of researchers had elaborately studied and reported BAs and their occurrences in human tissues [2], in plants [3], in foods and food products such as in fish and seafood [4,5], in meat products [6,7], in bean products [8], in fruits and vegetables [9,10], and in wine and alcoholic beverages [11,12]. Specifically, the most frequent BAs important in food analysis are the diamines, e.g. putrescine (PUT) and cadaverine (CAD); polyamines, e.g.

spermidine (SPD) and spermine (SPM); aromatic amines, e.g. tyramine (TYR); and heterocyclic amines, e.g. tryptamine (TRY) and histamine (HIS) [13]. Table 1 shows an example of these compounds. Generally, food products that depend on natural fermentation are more susceptible to biogenic amines as a result of a contaminating microflora exhibiting amino acid decarboxylase activity, such as cheese, soy bean products and alcoholic beverages[14].

There are many different analytical approaches advocated to extract BAs compounds from food products. Common methods applied for this application were: Liquid-liquid extraction method (LLE), Liquid-phase microextraction method (LPME), Solid-phase extraction method (SPE) [15] and Solid-phase microextraction method (SPME) [16].

Name	Abbreviation	Structure formula	Molecular weight		
Cadaverine	CAD	H <sub>2</sub> N NH <sub>2</sub>	175.1		
Histamine	HIS	N CH <sub>2</sub> CH <sub>2</sub> NH <sub>2</sub>	111.1		
Phenylethylamine	PEA	CH <sub>2</sub> CH <sub>2</sub> NH <sub>2</sub>	121.2		
Putrescine	PUT	H <sub>2</sub> NNH <sub>2</sub>	88.2		
Tryptamine	ТҮР	CH <sub>2</sub> CH <sub>2</sub> NH <sub>2</sub>	160.2		
Tyramine	TYR	HO-CH <sub>2</sub> CH <sub>2</sub> NH <sub>2</sub>	137.2		
Spermidine	SPD	H <sub>2</sub> N N NH <sub>2</sub>	145.3		

Table1. Structures of biogenic amines [8]

The abovementioned separation methods are usually used in conjunction with BA detection methods. The common separation and determination techniques used intensively for this purpose were known to be: Fluorescence-based methods [17], Capillary electrophoresis [18], Molecular/enzyme-based methods such as Polymerase Chain Reaction (PCR) method [19] and Enzyme-Linked Immunosorbent Assay (ELISA) method [20]. Chromatographic-based methods

such as the techniques of Thin Layer Chromatography (TLC) [21], High Performance Liquid Chromatography (HPLC) [22,23], Gas Chromatography (GC) [24]. Liquid Chromatography (LC) [25] and Ion-Exchange Chromatography (IC) [26].

Many methods have shown considerable variation in response to BA in terms of derivatization, suitability to BA compounds, detectability, recovery, selectivity, linearity, repeatability, reproducibility, sensitivity, accuracy, cost- and time-effectiveness, technical skills required, and instrumentation and preparation procedures. HPLC with pre- or post-column derivatisation is considered the most extensively reported method used in separation and quantification of BA due to its higher resolution, sensitivity, great versatility and simple treatment [27]. The present study comes profoundly to meet the aforementioned aspirations, by introducing hydrazone as a novel sorbent synthesized ligand. Since there is no work have been reported for the extraction of BAs using hydrazone compounds.

#### 2. Experimental

Analytical grade chemicals, reagents and solvents were used as received without additional purification. 18 m $\Omega$  ultrapure water was used throughout. A Perkin Elmer 200-series HPLC unit and an Ultraviolet-visible lambda 35 UV -vis spectrometer were used for separation and detection, respectively. Stuart Scientific flask shaker SFI (500 osc min<sup>-1</sup>) was used for the extraction. The chromatographic separation in the isocratic mode was carried out using a C<sub>18</sub> ODS Hypersil column (250 × 4.5 mm, 5 µm) at ambient temperature. The mobile phase was 60:25:15 (v/v/v) acetonitrile:water:methanol at a flow rate of 1.0 mL min<sup>-1</sup>; wavelength, 254 nm; and injection volume, 50 µL. The preparation of the ligand, the sorbent material and stock solution (1000 mg L<sup>-1</sup>) of a mixture of seven BAs (TRY, PEA, PUT, CAD, HIS, TYR and SPD) were carried out as previously reported [28,29].

### 2.1 Characterization of the ligand and sorbent

Because of the failure of the infrared technique to provide a conclusive evidence on the presence of the hydrazone compound in the gel matrix, which could be attributed to the too small amount of the ligand in the network, solid state UV-Visible analysis was carried out. A comparison between the free ligand and its corresponding sol-gel sorbent was carried out using solid state UV-Vis technique in the region from 200-800 nm. The free ligand showed red shift with maximum absorption ( $\lambda_{max}$ ) around 420 nm. The UV–Vis spectrum of the free ligand, blank and the hydrazone immobilized sorbent is shown in Fig. 1. The difference in the spectrum of the blank and sorbent is a strong evidence that indicates the successful incorporation of the hydrazone ligand into the sol-gel network. However, incorporating the ligand into the silica-gel causes a shift in thewavelength ( $\lambda_{max}$ ) of the free ligand ( $\Delta\lambda_{max}$ = -29). This could be due to the intermolecular interactions between the ligand and the silica which can change the characteristics of the absorption spectra.

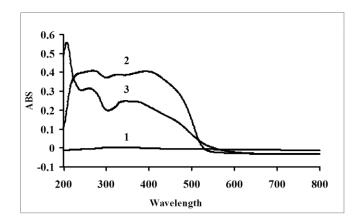
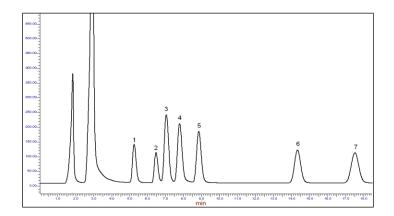


Figure 1. UV-Vis spectra of (1) blank sol-gel, (2) free ligand, and (3) sol-gel sorbent

# 2.2 Extraction method

Batch method of extraction was conducted. The sorbent (25 mg) was placed in a glass vial along with 1 mL BAs standard mixture and 4 mL of 0.1 M Tris buffer (pH 9). The mixture was shaken mechanically at room temperature for 15 min. After the equilibrium time, the mixture was filtered and the filtrate (1 mL) was derivatized previously reported [28]. The amount of the unextracted BAs left in the solution after the extraction was determined by reversed phase HPLC (Fig. 2). The extracted amount was calculated by the difference.



*Figure 2.* Chromatogram of BA (20 mg  $L^{-1}$  each) standard solution.

TRY (1); PEA (2); PUT (3); CAD (4); HIS (5); TYR (6); SPD (7).

# 3. Results and Discussion

The extraction was optimized by using 20 mg  $L^{-1}$  BAs standard solution. Important parameters that affects the extraction efficiency were investigated, i.e., sample pH, ligand concentration within the sorbent material, extraction time, and sorbent capacity. The extraction was also carried out using blank sorbent (i.e. does not contain the studied ligand) as control. The enhancement on the extraction behaviour between the blank and the ligand immobilized sorbents demonstrated the capability of hydrazone ligand in the extraction process (Fig. 3).

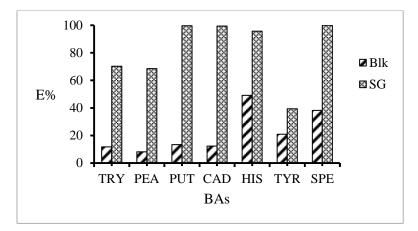


Figure 3: Extraction of BAs by the sol-gel matrix

# 3.1 Effect of contact time

The effect of contact time on the extraction efficiency was studied by shaking the BAs mixture with the sorbent material ranging from 5 to 60 min (Fig.4). Good extraction was observed after 5 min of contact time (%E >70 %) for all the studied BAs. However, maximum extraction was achieved after 15 min. Increasing the contact time further has no significant improvement of the %E. Therefore, 15 contact time was chosen for subsequent studies.

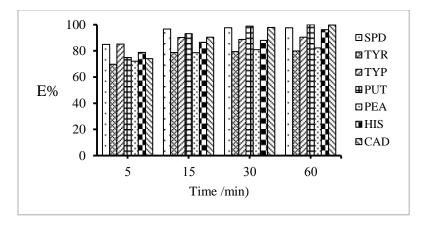


Figure 4: Effect of contact time on the extraction of BAs

# 3.2 Effect of ligand concentration in the sol-gel network

Sorbent materials containing different concentrations of ligand (0.85% - 6.8%)were prepared and their extraction efficiency was examined at the optimum pH 9 and extraction time of 15 min. It was observed that the highest extraction (%E >80) for all BAs was obtained when 0.85% ligand in the sorbent was used (Fig.5). Further increase of the ligand concentration does not seem to affect the extraction efficiency.

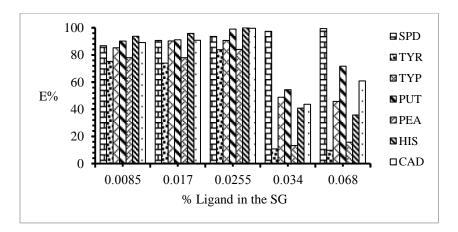


Figure 5 : Effect of ligand concentration on the extraction of BAs

# **3.3 Effect of BAs concentration**

Five different concentrations (50 - 400 mg  $L^{-1}$ ) of BAs were used to study the extraction efficiency and the capacity of the sorbent (Table2). It was found that the aliphatic BAs (PUT, CAD, and SPD) were quantitatively extracted (%E 100 %) up to 200 mg  $L^{-1}$ . At concentrations higher than 200 mg  $L^{-1}$  a decrease in the extraction of PUT and CAD was observed, while no decrease was found for SPD. The aromatic BAs were also poorly extracted than the aliphatic. The presence of the aromatic ring resulted in a decrease in the extraction efficiency due to steric effects. HIS was extracted more than the other aromatic BAs due to its smaller size.

BAs	Concentration (ppm)											
	5		50		100		200		400		500	
	BLK	SG	BLK	SG	BLK	SG	BLK	SG	BLK	SG	BLK	SG
TYR	0	100	64.10	77.68	47.18	67.36	38.23	56.13	34.42	46.66	32.46	45.67
PEA	0	0	0	24.01	4.94	31.02	9.19	29.37	13.43	29.92	14.92	31.19
PUT	54.15	100	29.47	100	30.18	100	21.08	100	21.67	61.61	24.05	51.51
CAD	50.19	100	35.76	100	34.29	100	24.31	97.42	24.32	52.97	27.15	44.54
HIS	100	100	74.75	99.39	48.36	99.85	36.02	68.89	30.17	44.29	31.43	42.06
TRY	0	51.30	13.37	61.41	14.07	32.43	11.51	30.61	18.29	28.10	33.06	29.77
SPD	57.19	100	48.46	100	44.99	100	34.03	100	42.94	98.95	53.17	95.92

Table 2 Effect of BAs concentration

BLK= blank sol-gel, SG= immobilized sol-gel (sorbent)

### 4. Conclusion

The studied hydrazone ligand shows preferential selectivity towards the aliphatic (PUT, CAD, SPD) and the heterocyclic (HIS) over the aromatic (TRY, PEA, TYR) BA. The high extraction efficiency of the sorbent was achieved due to the high affinity between the sol-gel sorbent that contained the hydrazone ligand and the target analyte. A significant advantage of

hydrazone compounds when compared with sorbents based on other ligands such as crown ethers is that the hydrazones can be easily synthesized using a one simple step at room temperature.

### References

- [1] Ekegren, T. and Gomes-Trolin, C. (2005). Determination of polyamines in human tissues by precolumnderivatization with 9-fluorenylmethyl chloroformate and high-performance liquid chromatography. *Analytical Biochemistry*, **338**, 179-185.
- [2] Walters, D. R. (2003). Polyamines and plant disease. *Phytochemistry*, **64**, 97-107.
- [3] Tsai, Y-H., Lin, C-Y., Chang, S-C., Chen, H-C., Kung, H-F., Wei, C-I. and Hwang, D-F. (2005). Occurrence of histamine-forming bacteria in salted mackerel in Taiwan. *Food Microbiology*, **22**, 461-467.
- [4] Auerswald, L., Morren, C. and Lopata, A.L. (2006). Histamine levels in seventeen species of fresh and processed South African seafood. *Food Chemistry*, **98**, 231-239.
- [5] Özogul, F. and Özogul, Y. (2006). Biogenic amine content and biogenic amine quality indices of sardines (*Sardinapilchardus*) stored in modified atmosphere packaging and vacuum packaging. *Food Chemistry*, **99**, 574-578.
- [6] Gençcelep, H., Kaban, G., Aksu, M.İ., Öz, F., and Kaya, M. (2008). Determination of biogenic amines in sucuk. *Food Control*, **19**, 868-872.
- [7] Kalač, P. and Krausová, P. (2005). A review of dietary polyamines: Formation, implications for growth and health and occurrence in foods. *Food Chemistry*, **90**, 219-230.
- [8] Chiacchierini, E., Restuccia, D. and Vinci, G. (2006). Evaluation of two different extraction methods for chromatographic determination of bioactive amines in tomato products. *Talanta*, **69**, 548-555.
- [9] Adão, R.C. and Glória, M.B.A. (2005) Bioactive amines and carbohydrate changes during ripening of 'Prata' banana (*Musa acuminate x M. balbisiana*). *Food Chemistry*, **90**, 705-711.
- [10] Valero, D., Martínez-Romero, D., and Serrano, M. (2002). The role of polyamines in the improvement of the shelf life of fruit. *Trends in Food Science & Technology*, **13**, 228-234.
- [11] Soleas, G. J., Carey, M. and Goldberg, D. M. (1999). Method development and cultivar-related differences of nine biogenic amines in Ontario wines. *Food Chemistry*, **64**, 49-58.
- [12] Vidal-Carou, M. C., Lahoz-Portolés, F., Bover-Cid, S. and Mariné-Font, A. (2003). Ion-pair highperformance liquid chromatographic determination of biogenic amines and polyamines in wine and other alcoholic beverages. *Journal of Chromatography A*, **998**, 235-241.
- [13] Stratton, J. E., Hutkins, R. W. and Taylor, S. L. (1991). Biogenic amines in Chinese and other fermented foods: A review. *Journal of Food Protection*, **54**, 460-470.
- [14] Santos, M. H. S. (1996). Biogenic amines: their importance in foods. International Journal of Food Microbiology, 29, 213-231.
- [15] Valero, D., Martínez-Romero, D., and Serrano, M. (2002). The role of polyamines in the improvement of the shelf life of fruit. *Trends in Food Science & Technology*, **13**, 228-234.
- [16] Anli, R.E., Vural, N., Yilmaz, S. and Vural, Y.H. (2004). The determination of biogenic amines in Turkish red wines. Journal of Food Composition and Analysis, 17, 53-62.
- [17] Soleas, G. J., Carey, M. and Goldberg, D. M. (1999). Method development and cultivar-related differences of nine biogenic amines in Ontario wines. Food Chemistry, 64, 49-58.
- [18] Sano, M. and Nishino, I. (2007). Assay for spermidine synthase activity by micellar

electrokinetic chromatography with laser-induced fluorescence detection. *Journal of Chromatography B*, **845**, 80-83.

- [19] Kvasnička, F. and Voldřich, M. (2006) Determination of biogenic amines by capillary zone electrophoresis with conductometric detection. *Journal of Chromatography A*, **1103**, 145-149.
- [20] Landete, J. M., de las Rivas, B., Marcobal, A. and Muñoz, R. (2007). Molecular methods for the detection of biogenic amine-producing bacteria on foods. *International Journal of Food Microbiology*, **117**, 258-269.
- [21] Marcobal, A., Polo, M. C., Martín-Álvarez, P. J. and Moreno-Arribas, M. V. (2005). Biogemic amine content of red Spanish wines: comparison of a direct ELISA and an HPLC method for the determination of histamine in wines. *Food Research International*, **38**, 387-394.
- [22] Shakila, R. J., Vasundhara, T. S. and Kumudavally, K. V. (2001). A comparison of the TLCdesitometry and HPLC method for the determination of biogenic amines in fish and fishery products. *Food Chemistry*, **75**, 255-259.
- [23] Soufleros, E. H., Bouloumpasi, E., Zotou, A. and Loukou, Z. (2007). Determination of biogenic amines in Greek wines by HPLC and ultraviolet detection after dansylation and examination of factors affecting their presence and concentration. *Food Chemistry*, **101**, 704-716.
- [24] Yongmei, L., Xin, L., Xiaohong, C., Mei, J., Chao, L. and Mingsheng, D. (2007). A survey of biogenic amines in Chinese rice wines. *Food Chemistry*, **100**, 1424-1428.
- [25] Hwang, D-F., Chang, S-H., Shiua, C-Y., and Chai, T-J. (1997) High-performance liquid chromatographic deteremination of biogenic amines in fish implicated in food poisoning. *Journal of Chromatography B*, **396**, 23-30.
- [26] Gianotti, V., Chiuminatto, U., Mazzucco, E., Gosetti, F., Bottaro, M., Franscarolo, P. and Gennaro, M. C. (2008). A new hydrophilic interaction liquid chromatography tandem mass spectrometry method for the simultaneous determination of seven biogenic amines in cheese. *Journal of Chromatography A*, **1185**, 296-300.
- [27] Draisci, R., Giannetti, L., Boria, P., Lucentini, L., Palleschi, L. and Cavalli, S. (1998) Improved ion chromatography – integrated pulsed amperometric detection method for the evaluation of biogenic amines in food of vegetable or animal origin and in fermented foods. *Journal of Chromatography A*, **798**, 109-116.
- [28] Abdassalam Abdelhafiz. Tameem, B. Saad, A. Makahleh, A. Salhin, M. I. Saleh, Talanta 82(2010) 1385.
- [29] Chanbasha Basheer, Weishan Wong, Ahmad Makahleh, Abdassalam Abdelhafiz Tameem, Abdussalam Salhin, Bahruddin Saad, and Hian Kee Lee, Journal of Chromatography A.1218(2011) 4332-4339.