# Isoenzyme diversity for peroxidase among Libyan thyme (*Thymus capitatus* (L.) Hoffm & Link.) Populations

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**ABSTRACT:** Thyme (*Thymus capitatus* (L.) Hoffm & Link) is an endemic wild plant in southern parts of Al-Jabal Al-Akhdar region, Libya. There are five phenotypes (populations or accessions) of thyme that show flower color polymorphism. Difference in flower color among the five phenotypes is due to genetic variation. This study was conducted to determine the pattern of peroxidase isoenzymes in thyme which growing in south parts of Al-Jabal Al-Akhdar . Laves samples of the five *Thymus capitatus* (L.) Hoffm & Link genotypes (populations) were evaluated for peroxidase isoenzyme system by horizontal starch gel electrophoresis. All five phenotypes had different number and relative mobility of electrophoretic bands. This variability enabled the five phenotypes of *Thymus capitatus* (L.) Hoffm & Link to be distinguished.

Key words: electrophoresis, genetic diversity, endemic species, peroxidase, *Thymus capitatus* (L.) Hoffm & Link.

#### INTRODUCTION

Systems of classification and identification based on quantitative and/or qualitative morphological characters were used by researchers to study genetic diversity (Trujillo *et al.*, 1995; Ali *et al*, 2019). Although these methods were effective, they presented practical drawbacks due to effect of environmental fluctuations on expression of most morphological traits (Degani *et al*, 1995; Ali and Mustafa, 2019). The use of biochemical markers such as isoenzymes overcome these problems since they are little affected by the environment and can easily be detected in a variety of tissues by relatively simple, rapid and inexpensive procedures (Weeden and Lamb, 1985; Obara-Okeyo *et al.*, 1997).

The fact that different molecular forms of enzyme which catalyze the same reaction (isozymes) can occur in the same organism is providing to be not only an especially valuable aid in many biological studies, but it is also providing a new and exciting perspective for the interpretation of a number of central problems to modern biological thoughts such as cellular differentiation, genetics and evolution (El-Wakil *et al*, 2000).

Horizontal starch gel electrophoresis, which uses the migration of proteins in an electric field to detect small differences in their charge and shape, has proved to be useful tool for identification of genetic diversity (Cyrus and Cummins, 1992; Ali, 1999). Isoenzymes analysis by horizontal starch gel electrophoresis can help validate plant pedigree, phylogeny, polyploidy, and mating system and help distinguish among populations (Peirce and Brewbaker, 1973).

Thyme (*Thymus*) is a genus containing about 350 species of aromatic perennial herbs and sub-shrubs to 40 cm tall, belong to the family Lamiaceae. This family is distributed throughout the arid, temperate and cold regions including Europe, North Africa and Asia. It is in leaf all year, flowering from July to September (Gruenwald *et al.*, 2004).

Thyme (*Thymus capitatus* (L.) Hoffm & Link) is a perennial woody plant, 15 cm tall, with abundant stoloniferous branches. The inflorescences are composed of whorls of small, zygomorphic flowers. The pollination in *T.capitatus* is entomophilous, as in most *Thymus* 

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species. The main pollinators are *Apis mellifera*. *T. capitatus* is gynodioecious, as for most species of *Thymus* (Ali and Mustafa, 2019). *Thymus capitatus* is an endemic wild plant in southern parts of Al-Jabal Al-Akhdar, Libya. Thyme growing in southern parts of Al-Jabal Al-Akhdar showed flower color polymorphism which results five different phenotypes: white- flowered, dotted white-flowered, purple-flowered, violet-flowered and mosaic-flowered populations (Ali and Mustafa, 2019).

Isozyme analysis which has been successfully applied to identify several plant species, offers a possible alternative method for identification of different thyme populations. The aim of the present study was to identify thyme (*Thymus capitatus*) populations growing in south parts of Al-Jabal Al-Akhdar region, Libya by using isoenozyme analysis (peroxidase).

#### MATERIAL AND METHODS

The study was carried out to investigate the biochemical genetic markers (peroxidase isozyme) in natural populations of *Thymus capitatus* (L.) Hoffm & Link which growing in southern parts of Al-Jabal Al-Akhdar, Libya. *Thymus capitatus* (L.) Hoffm & Link shows a stable and dramatic flower-color polymorphism (Ali and Mustafa, 2019). There are five patterns: white-flowered, dotted white-flowered, purple-flowered, violet-flowered and mosaic-flowered plants present in natural populations of *Thymus capitatus* (L.) Hoffm & Link.

Peroxidase isozyme system were resolved by horizontal gel electrophoresis, using 12% hydrolyzed potato starch from Sigma. Leaf samples from fifteen plants for each population were homogenized using 0.1 ml of 0.23M Tris-citrate buffer ( pH 8.0) in a prechilled pestle and mortar. The homogenate was absorbed onto a small rectangle (4x2mm) of filter paper ( Whatman No. 1). These strips of filter paper were placed on the origin line of agar gel plates, for about 30 min. at 4°C. The filter paper strips were removed, then, an electrical constant current of 15-18 V/ cm was applied for 90 min. at 4°C using 0.23 M Tris-citrate buffer (pH 8.0), as an electrode buffer. After separation of the peroxidase isozymes, the gel plates were incubated at 4°C for 10 min. and were stained with peroxidase staining solution.

Staining solution for peroxidase was consists of 100ml. of 0.01Msodium acetate buffer (pH5.0), containing 0.1g benzidine and 0.5% hydrogen peroxide ( $H_2O_2$ ), which is added immediately before staining.

#### RESULTS

Staining for peroxidase (PX) after horizontal starch gel electrophoresis resulted in obvious phenotypic differences among natural populations or accessions of Libyan thyme (*Thymus capitatus* (L.) Hoffm & Link) when leaf tissue was used. The zymogram produced from isozymatic analysis clearly indicated that there was considerable variation among populations (accessions) of *Thymus capitatus* (L.) Hoffm & Link for peroxidase (PX). All the bands were consistent and repeatable in all different accessions (Fig. 1).

Two polymorphic regions of peroxidase (PX) activity were observed on the gel. One moved cathodally and the other moved anodally. The zones of activity (bands) for peroxidase were numbered according to their proximity to the original line (O.L.). The intensity of the bands was illustrated with different strips: black, gray and white, which indicate high, medium and low activities of peroxidase, respectively (Fig. 1). This figure was more illustrated by figure 2.

Polymorphism was observed in peroxidase system for all natural populations of thyme (*Thymus capitatus* (L.) Hoffm & Link) growing in southern parts of Al-Jabal Al-Akhdar region, Libya. The present data identified five isoenzymes of peroxidase expressed as different banding patterns (Fig.1, 2). Three bands were anodal ( $PX_1$  to  $PX_3$ ) and one band was cathodal ( $PX_4$ ).

Results of the present study showed that populations of Libyan thyme had different relative mobility (Rf) of electrophoretic bands. PX zymogram gave five different banding patterns composed of one to four bands, with the fastest band (PX1) at 0.5 Rf and the slowest band (PX3) at 0.2 Rf. The values of relative mobility for PX2 and PX4 were 0.38 and 0.25, respectively (Table 1).

Except violet-flowered population, all populations of *Thymus capitatus* (L.) Hoffm & Link in the present study had PX3 band (Fig. 2), while only two populations (purple-flowered and mosaic-flowered accessions) carried a fast band (PX1). Moreover, the PX4 band was absent in white-flowered and violet-flowered plants. On other hand, the PX2 band was present in all populations except white-flowered and mosaic-flowered populations (Table 1).



Fig (1): Photograph of banding patterns for peroxidase in *Thymus capitatus* (L.) Hoffm & Link populations.



Fig (2): Diagrammatic representation of banding patterns and populations observed in *Thymus capitatus* (L.) Hoffm & Link for peroxidase.



Table (1): Activity and Relative mobility of banding patterns for peroxidase in Thymuscapitatus (L.) Hoffm & Link populations.

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			Anodal direction	l	Cathode direction
	Band	Px1	Px2	Px3	Px4
Relative mobility		0.50	0.38	0.20	0.25
Populations	White-flowered			В	
	Dotted white-flowered		В	В	C
	Purple-flowered	А	А	В	C
	Violet-flowered		C		
	Mosaic-flowered	С		В	C

A= High activity B = Medium activity C = Low activity

## DISCUSSION

Plant peroxidases are contained mainly in cell walls and such localization results in their exhibiting activity already at early stages of defense reactions of the plant to pathogen attacks (Rautela and Payne 1970; Van-Geyt,1986; Cook *et al.*, 1995; Harrison *et al.*, 1995; Curtis *et al.*, 1995; Thordal-Christensen *et al.*, 1992). The reactions are revealed by accelerated lignification (Vance *et al.*, 1980; Gaspar *et al.*, 1982; Walter, 1992), and suberisation (Goldberg *et al.*, 1981), as well as production of anti-microbial radicals (Peng and Kuc, 1992; Kobayashi *et al.*, 1994). Besides, peroxidases can produce oxidized phenolics or other substances toxic for invading pathogens (Gaspar *et al.*, 1991). Peroxidases are ubiquitous in plants. They have become standard isoenozyme markers due to their easy detection. In addition, the enzymes are extremely stable (Rasmussen and Kerby,1993; Kolodziejczak and Krzakowa, 2003).

In this work no inheritance study was done for making the different between allozyme and isoenzyme bands. So the genetic differences among different populations were mainly depended on the difference between populations in their banding patterns as a whole.

This study shows that isozymes can be used for identification of Libyan thyme populations at the vegetative stage. This should be useful to germplasm repositories for early detection of mislabeled plants in natural populations of thyme and therefore help maintain the purity of thyme stocks. Moreover, the observed variability can be useful in germplasm evaluation. Also, precise identification of thyme natural populations by isozymes can be useful for patent purposes. These observations agree with those of others (Loapez-Pujol *et al*, 2004; Bartošova *et al.*, 2005). Although morphological and physiological markers occasionally may be sufficient for identification, isoenzyme is essential when these markers are not available (Degani and Blumenfeld, 1986).

The present investigation showed that natural populations of Libyan thyme can be distinguished on the basis of peroxidase isozyme system making the identification to be economical and applicable in laboratories. Peroxidase system seemed to be useful in discriminating among thyme populations because of a variety of banding patterns. These findings are in agreement with findings of many previous studies (Byrne and Littleton, 1988; Hannan and Orick, 2000; Stoyanov and Stoyanova, 2007).

The current study indicated that peroxidases of Libyan thyme (*Thymus capitatus* (L.) Hoffm & Link) also exhibit cathodal and anodal migration. These results agree with many researches which reported that most plants contain peroxidases of cathodal and anodal migration. Peroxidases of cathodal and anodal migration were detected in different, taxonomically very distant groups of plants, e.g. in bryophytes (Krzakowa, 1991), grasses

(Felder, 1976; Krzakowa, 1996), peanut (Wan and Vanhusystee, 1993), and in radish (Kim and Park, 1996).

Furthermore, Peroxidase profiles have been used as taxonomic markers and measures of inter-population variability in the *Sphagnum subsecundum* complex, where the lack of morphological discontinuities renders the separation of taxa based only on these features controversial and pointless (Abe and Tsuda, 1987; Abe *et al.*, 1993; Krzakowa and Melosik, 2000).

The Libyan thyme populations that examined in this study exhibited substantial isoenzymic variability that proved useful in identification and verification. This variability is a reflection of diverse and complex species background and its out-crossing nature. These results showed that isoenzyme analysis not only provides genetic markers for identification purpose but is also a procedure for analyzing the origin of species (Hauagge *et al*, 1987; Byrne and Littleton, 1988; Volis *et al.*, 2002; Kolodziejczak and Krzakowa, 2003).

These results are expected because these populations are reproduced by seeds. Therefore, the genetic variability among thyme populations is mostly due to genetic segregation produced from sexual reproduction. These findings were confirmed by several studies on different plant species (El-Wakil *et al.*, 2000; Loapez-Pujol *et al.*, 2004; Talukdar, 2010).

المستخلص: الزعتر (Thymus capitatus) نبات بري متوطن في الجزء الشرقي من ليبيا. يوجد خمسة أشكال مظهرية لنبات الزعتر البري تنمو بمنطقة جنوب الجبل الأخضر. الأشكال المظهرية الخمسة لهذا النبات أظهرت تعدد أنماط بالنسبة لصفة لون الأزهار. الاختلاف في لون الأزهار بين الأشكال المظهرية الخمسة يرجع إلى التباين الوراثي. هذه الدراسة أجريت لغرض تحديد مشابحات أنزيم peroxidase في نبات الزعتر المنتشر بمنطقة جنوب الجبل الأخضر. عينات الأوراق للأشكال المظهرية الخمسة جمعت لتحديد غط مشابحات أنزيم peroxidase في نبات الزعتر الكهربائية الأفقي بملام النشا. الأشكال المظهرية الخمسة لنبات الزعتر أظهرت اختلاف في العدد و الحركة النسبية لحزم الاختلاف يجعل من الممكن التمييز بين الأشكال المظهرية الخمسة لنبات الزعتر المهرت اختلاف في العدد و الحركة النسبية لحزم التفريد الكهربائي.

## **REFERENCES:**

1. Abe, J., Guang, P. and Shimamoto, Y. (1993). Linkage maps for nine isozyme and four marker loci in sugarbeet (*Beta vulgaris* L.). Euphytica, 66: 117-126.

2. Abe, J. and Tsuda, Ch. (1987). Genetic analysis for isozyme variation in the section *Vulgares*, Genus *Beta*. Japan. J. Breed. 37: 253-261.

**3.** Ali, E. S. (1999). Genetical and cytological studies on some olive cultivars (*olea europaea* L.) in Egypt. M. Sc. Thesis. University of Alexandria, Egypt.

4. Ali, E. S. and Mustafa, H. M. (2019). Polymorphism in thyme (*Thymus capitatus*) at southern region of El-Jabal El-Akhdar, Libya. Bayan Journal of Sciences, 1(4): 47-62.

5. Ali, E. S., Mustafa, H. M. and Blkasem, K. O.(2019). Morphological variation of Libyan carob (*Ceratonia siliqua* L.). Al-Mukhtar Journal of Sciences, 34(2): 126-133.

6. Bartošova Z, Obert B, Takac T, Kormutak A, Pretova A (2005). Using enzyme polymorphism to identify the gametic origin of flax regenerants. Acta Biologica Cracoviensia series Botanica, 47(1): 173-178.

7. Byrne, D. and Littleton, T. (1988). Electrophoretic characterization of diploid plums of the Southeastern United States. J. Amer. Soc. Hort. Sci. 113(6); 918-924.

8. Cook, D., Dreyer, D., Bomet, D., Howell, M., Nony, E. and Vanden-Bosch, K. (1995). Transient induction of a peroxidase gene in *Medicago truncatula* precedes infection by *Rhizobium meliloti*. Plant Cell. 7: 43-45.

9. Curtis, M.D., Nourse, J.P. and Manners, J.M. (1995). Nucleotide sequence of a cationic peroxidase gene from the tropical forage legume *Stylosanthes humilis*. Plant Physiol. 108: 1303-1304.

10. Cyrus, S. and Cummins, J. (1992). Distinguishing apple rootstocks by isozyme banding patterns. HortiScience, 27(7): 829-831.

11. Degani, C., Beiles, R., El-Batsri, M., Goren, M. and Gazit, S. (1995). Identifying Lychee cultivars by isozyme analysis. J. Amer. Soc. Hort. Sci., 120(2): 307-312.

12. Degani, C. and Blumenfeld, A. (1986). The use of isozyme analysis for differentiation between loquat cultivars. J. Amer. Soc. Hort. Sci. 21 (6): 1457-1458.

13. El-Wakil, H. E., Harhash, M. M., Khaled, A. S. and Ali, E. S. (2000). Identification of some olive cultivars by isozyme analysis and fruit properties. Adv. Agric. Res., 5(1): 1131-1147.

14. **Felder, M. R. (1976).** Genetic control of four cathodal peroxidase isozymes in barley. J. Hered., 67: 39-42.

15. Gaspar, T.H., Penel, C., Hageged, D. and Greppin, H. (1991). Peroxidases in plant growth, differentiation and development. In: Biochemical, molecular and physiological aspects 60 M. Ko<sup>3</sup>odziejczak, M. Krzakowa of plant peroxidases (J. Lobarzewski, H. Greppin, C. Penel, Th. Gaspar, eds.). University of Geneva, Switzerland: 249-280.

16. Gaspar, T.H., Penel, C., Thorpe,T. and Greppin, H.(1982). Peroxidases 1970-1980. Asurvey of their biochemical and physiological roles in higher plants. Universite de Geneve, Geneve, Switzerland: 89-121.

17. Goldberg, R., Catesson, A and Szaninsky, Y.(1981). Histochemical and biochemical characteristics of peroxidase involved in lignification processes of poplar. In: Cell Wall's 81(D.G. Robinson, H. Quader, eds.). Proc. Zol. Cell Wall Meeting:251-260.

18. **Gruenwald, J., Brendler, T. and Jaenicke, C. (2004).** PDR for Herbal Medicines, 3rd edition. Montvale (NJ): Thompson PDR, pp.815-816.

19. Hannan, G., Orick, M. (2000). Isozyme diversity in Iris cristata and the threatened glacial endemic I. lacustris (Iridaceae). American Journal of Botany 87: 293-301.

20. Harrison, S., Curtis, M., McIntyre, C. Maclean, D. and Manners, J. (1995). Differential expression of peroxidase isogenes during the early stages of infection of the tropical forage legume *Stylasanthes humilis* by *Colletotrichum gloeosporioides*. Molecular Plant Microbe Interactions, 8:398-406.

21. Hauagge, R., Kester, D., Arulsekar, D. Parfit, D. and Liu, L. (1987). Isozyme variation among California almond cultivars: II. Cultivar characterization and origins. J. Amer. Soc. Hort. Sci. 112: 693-698.

22. **Kim, S. S. and Park, J. H. (1996).** Genomic sequence and heterologous expression of a cationic isoperoxidase from radish. In: Plant peroxidases biochemistry and physiology (C. Obinger, K. Burner, R. Ebermann, C. Penel, H.Greppin, eds.). University of Geneva: 190-197.

23. Kobayashi, A., Koguchi, Y., Kanzaki, H., Kajiyama, S., Kawazu, K. (1994). A new type of antimicrobial phenolic produced by plant peroxidase and its possible role in the chemical defense systems against plant pathogens. Z. Naturforsch. 49: 411-414.

24. Kolodziejczak, M. and Krzakowa, M. (2003). Variability of cathodic peroxidases in sugar beet (*Beta vulgaris* L.) cultivars J. Appl. Genet., 44(1): 55-62.

25. **Krzakowa, M. (1991).** Peroxidases in genetic and taxonomic investigations. In: Biochemical, molecular and physiological aspects of plant peroxidases (J. Lobarzewski, H. Greppin, C. Penel, Th. Gaspar, eds.).UMCS and University of Geneva:15-20.

26. **Krzakowa, M. (1996).** Genetic diversity of *Phragmites australis* (Cav.)Trin. ex Stued revealed by electrophactically detected differences in peroxidases. In: Plant peroxidases: biochemistry and physiology (C. Obinger, U. Burner, C. Penel, H. Greppin, eds.). University of Geneva: 184-189.

27. Krzakowa, M. and Melosik, I. (2000). Taxonomic value of electrophoretically detected peroxidase patterns in four *Sphagnum* species (section *Subsecunda*, *Bryophyta*). Pl. Peroxid. Newslett., 14: 21-27.

28. Loapez-Pujol, J., Bosch, M., Simon, J. and Blanche, C. (2004). Allozyme diversity in the tetrapolid endemic *Thymus loscosii* (*Lamiaceae*). Annals of Botany 93: 323-332

29. **Obara-Okeyo, P., Fujii, K. and Kako, S. (1997).** Enzyme polymorphism in Cymbidium orchid cultivars and inheritance of Leucine aminopeptidase. HortiScience, 32(7): 1267-1271.

30. Peirce, L. C. and Brewbaker, J. L. (1973). Applications of isozyme analysis in horticultural science. HortScience, 8:17-22.

31. **Peng, M. and Kuc, J. (1992).** Peroxidase – generated hydrogen peroxide as a source of antifungal activity in vitro and on tobacco leaf disks. Phytopathology, 82: 696-699.

32. **Rasmussen, S. K. and Kerby, K. (1993).** Chromosomal localization of plant peroxidase genes In: Plant peroxidases: biochemistry and physiology (K.G. Welinder, S.K. Rasmussen, C. Penel, H. Greppin, eds.). University of Geneva 207-212.

33. Rautela, G. S. and Payne, M. G. (1970). The relationship of peroxidase and ortodiphenol oxidase to resistance of sugar beets to *Cercospora* leat spot. Phytopathol., 60: 238-245.

34. **Stoyanov, K. and Stoyanova, S. (2007).** Polymorphism in peroxidase isozymes detected in *Phelipanche (Orobanchaceae)* from Bulgaria. Phytologia Balcanica, 13 (2): 209–212.

35. **Talukdar**, **D.** (2010). Allozyme variations in leaf esterase and root peroxidase isozymes and linkage with dwarfing genes in induced dwarf mutants of grass pea (*Lathyrus sativus* L.). International Journal of Genetics and Molecular Biology, 2(6): 112-120.

36. Thordal-Christensen, H., Brand, J., Cho, B. H., Ramussen, S. K., Gregersen, P.L., Smedegaard-Peterson V. and Collinge, D. B. (1992). cDNA cloning and characterization of two barley peroxidase transcripts induced differentially by the powdery mildew fungus *Erisiphe graminis*. Physiol. Mol. Plant Pathol. 40: 395-409.

37. Trujillo, I, Rallo, L. and Arus, P. (1995). Identifying olive cultivars by isozyme analysis. J. Amer. Soc. Hort. Sci., 120:318-324.

38. Vance, C. P., Kirk T. K. and Sherwood, R. T. (1980). Lignification as a mechanism of disease resistance. Ann. Rev. Phylopathol., 18: 259-288.

39. Van Geyt, J. (1986). The use of an isozyme marker system in sugar-beet virus infection. In: Active defense mechanisms in plants (R.K.S. Wood, ed.). Plenum Publ. New York, 247-274.

**40.** Volis, S., Mendlinger, S., Turuspekov, Y. and Esnazarov, U. (2002). Phenotypic and allozyme variation in Mediterranean and desert populations of wild barley, Hordeum spontaneum koch. Evolution, 56(7): 1403–1415.

41. **Walter, M. H. (1992).** Regulation of lignification in defense. In: Gene involved in plant defense (T. Booler, F. Meins, eds.). Springer-Verlag. New York: 327-352.

42. Wan, L. and Vanhuystee, B. (1993). A study on glycolization of cationic peanut peroxidase. Biophys. Res. Commun., 197: 1398-1405.

43. Weeden, N. F. and Lamb, R. C. (1985). Identification of apple cultivars by isozyme phenotypes. J. Amer. Soc. Hort. Sci., 110:509-515.