Morphological study of *Serrasentis sagittifer* parasitic infecting the, fishes *Mullus barbatus* (Mullidae) from A-khoms coast, Libya

*.Abdusalam saleh Abusdel **.Atia Ramadan Elkilany *.Milud asaid alshaafai

*. Najwa abdalgader algaw

Abstract: A total of 120 marine fish as (*Mullus barbatus*) were collected from The Mediterranean Sea during the period from June 2021 to December 2021; they were dissected and examined to identify parasitic acanthocephali. Only 29 samples (24.16%) were infected with *Serrasentis sagittifer* from the family Rhadinorhynchidae.

Infested fishes illustrate no pathognomonic signs except abdominal distension, whereas the infection was recorded in the intestines, appendix and external surfaces of some internal organs of infected fish. Seasonally, the infection frequency increased to 55.17% during the summer season and decreased to 31.03 and 13.80 % during Autumn and winter respectively. Light microscopic and scanning electron microscopic examination revealed that the adult worm was elongated (wide anterior and narrow posterior ends) and measured 5.8-9.7 $(6.9\pm0.2)\times0.52-0.69$ (0.58 ± 0.02) mm for male and 10.1-11.3 $(10)\times0.68-0.83$ (0.72 ± 0.01) mm for female. The Proboscis was long of 0.89-1.3 mm (1.1 ± 0.2) for male and 1.11-1.19 mm (1.17 ± 0.02) for female. It was covered with many identical thorns arranged lengthwise as 9-11 Rows, each equipped with 15-18 spines. The body was supported by numerous vertebral ridges (16-20). Therefore, according to the morphological ⁱⁱⁱdocuments, the current parasite is classified as *Serrasentis sagittifer* and belongs to the class Palaeacanthocephala and the family Rhadinorhynchidae with a new host record from the Mediterranean Sea bream *Mullus barbtus*.

Keywords: Acanthocephalans; Serrasentis sagittifer; Mullus barbtus; alkuoms Coast; Libya; marine fish.

دراسة مورفولوجية لطفيلي serrasentis sagittifer الذي يصيب أسماك serrasentis sagittifer من ساحل الخمس بليبيا

المستخلص: أجريت هذه الدراسة على 120 سمكة من أسماك التريليا الحمراء (Mullus barbatus) من منطقة الخمس ليبيا لساحل الجنوبي للبحر المستخلص: أجريت هذه الدراسة على 2021 إلى ديسمبر 2021؛ تم تشريح وفحص العينات للبحث عن الديدان مشوكة الرأس الطفيلية. حيث تم الأبيض المتور على 29 عينة (24.16٪) فقط مصابة بشكل طبيعي بـ Serrasentis sagittifer التي تنتمي إلى عائلة العثور على 29 عينة (24.16٪) فقط مصابة بشكل طبيعي بـ Rhadinorhynchidae الإصابة في الأمعاء البوابية والأمطح الخارجية لبعض الأعضاء الداخلية للأسماك المصابة. موسمياً، ارتفع معدل انتشار العدوى إلى 55.17٪ خلال فصل الصيف وانخفض إلى 31.03 و13.80٪ خلال الخريف والشتاء على التوالي.

9.7-5.8 كشف الفحص المجهري للضوء والمسح الإلكتروني أن الدودة البالغة كانت ممدودة (مع نحايات خلفية أمامية وضيقة عريضة) وقياسها 9.7-5.8 (0.00 ± 0.70) $0.83-0.68 \times (0.2\pm0.8)$ (0.00 ± 0.70) مم للذكور و 0.01 ± 0.70 (0.00 ± 0.70) مم للذكور و 0.01 ± 0.70 ملم للإناث. كان الخرطوم طويلًا واسطوانيًا بطول 0.80-0.51 ملم (0.01 ± 0.70) للذكور و 0.01 ± 0.70 ملم الأشواك المنتظمة المرتبة طوليًا في 0.02 ± 0.70 منها ب 0.02 ± 0.70 عمودًا. كانت الأشواك مثلثة الشكل، على شكل

_

^{*} Biology Department Fac. Sciences El mergib Univ., Libya asabusdel@elmergib.edu.ly

^{**} Biology Department Fac. Education El mergib Univ., Libya arelkilany@elmergib.edu.ly

^{*} Biology Department Fac. Sciences El mergib Univ., Libya milud@elmergib.edu.ly

^{*} Biology Department Fac. Sciences El mergib Univ., Libya naalgaw@elmergib.edu.lu

سهم، قوية؛ انخفض حجمها من القمة إلى قاعدة خرطوم. يتبع الخرطوم منطقة عنق قصيرة غير شوكية متبوعة ومدعومًا بأمشاط متعددة من الأشواك (20-16) على سطحه البطني. تدعمها المعطيات المورفولوجية ووجود جذع العمود الفقري مرتبة ضمن صفوف (شبيهة بالمشط) ووجود أربع غدد أسمنتية في الذكور. لذلك، وفقًا لسجلات الشكل المورفولوجي، يتم تصنيف الطفيل الحالي على أنه Serrasentis sagittifer الذي ينتمي إلى فئة Palaeacanthocephala وعائلة Phadinorhynchidae مع سجل مضيف جديد من اسماك Mullus barbtus في البحر الأبيض المتوسط لبيا.

Introduction

The Mediterranean Sea has a very rich and varied fish fauna. It is one of the major centers of global marine biodiversity, Marine fishes are considered as one of the most important sources of animal protein in Libya.

Knowledge of fish parasites is of particular interest, not only for fish health but also for understanding ecological problems. The parasites play important role in the lives of their hosts (**Dogiel, 1964**), and they have been proposed as excellent indicators of the biodiversity both on host species and at the ecosystem level (**Chambers and Dick, 2005**).

Acanthocephalans are a group of Endoparasites helminthe found commonly in both marine and freshwater fishes. They are characterized by complex life cycle including arthropods as intermediate hosts and vertebrates as definitive or paratenic hosts. They cause pathological conditions in many fish (Nickol, 2006). Attachment by the armed the proboscis may cause mechanical damage which affects the architecture of the intestinal tissues. Heavy infections may cause obstruction of the intestines and invasion or migration of the helmet parasites to uncommon locations (Sanil et al., 2010 and Sakthivel et al., 2014).

Acanthocephalan fish parasites live as adults in the intestine or as larvae in fish tissue. All acanthocephalans use arthropods as intermediate hosts and vertebrates as definitive hosts. Acanthocephalan larvae that develop into adults only when ingested by appropriate definitive hosts. (**Tingbao and Xianghua, 2001**).

Materials and Methods

120 fish specimens of *Mullus barbatus* (Mullidae) were collected from the Mediterranean Sea, Libya during the period from July to December 2021. helminths were isolated from the Intestine, pyloric ceca, body cavity, mesenteries and external surfaces of internal organs of the fishes transfer done to a clean 0.9 % saline solution by using a brush or small pipettes and wash several times to remove any mucous or debris which is usually adhere to body surface. (**Gibson, 1985**), Then fixed in 10% formalin.

After fixation, the collected samples were washed in distilled water for 15 minutes. This should be carried out to remove the excess stain by placing the stained helminth into a dilute solution of acid alcohol (0.5 ml in 1000 ml alcohol). Following the upper procedure, it was followed by dehydration in an ascending series of ethanol, 10%,20%,30%,40, 50%,60%, 70%,90%,95% and absolute ethanol, leaving the parasites for 2-5 minutes in each grade. The samples were then cleared in xylene mounted in Canada balsam, covered with cover glass and left to dry in. The prevalence morphometric measurements (**Bush et al 1997**) minimum and maximum values were given, followed in parentheses by the arithmetic mean \pm SD.

For scanning electron microscopy, the samples were fixed in 3% buffered glutaraldehyde, and dehydrated in an ascending alcohol series. After passing through an ascending series of the Genosolv-D, they were processed "Bomer-900" with Freon 13 and sputter coated with gold–palladium in a Technics Hummer V and examined with an Etec Autoscan at 25 kV

Result

In present study, an acanthocephalan parasite was recovered from the intestine of *Mullus barbatus* captured from marine fishes the Mediterranean Sea in al- khoms Libyan coasts. The parasite was observed attached to the wall of the host intestine by an armed proboscis equipped by recurve hooks.

Clinical findings

Infested fishes showed no pathognomonic signs except abdominal distension and Infection was reported in the intestine, pylorus of the appendix, and on the external surfaces of some internal organs of infected fish. Twenty nine out of 120 fish specimens (24.16%) infected, seasonally the it has risen of infection increased to 55.17 % during summer season and decreased to 31.03 and 13.80 % during Autumn and winter respectively. The average parasite presence was 3 to 7 parasites per infected fish. The recovered helminthe were creamy white, elongated with narrow posterior end.

scanning electron and Light microscopy showed that the parasite had distinctive rows of spines (combs) on the ventral surface and measured 5.8 to $9.7~(6.9\pm0.2)\times0.52-0.69~(0.58\pm0.02)$ mm for male While female and $10.1-11.3~(10)\times0.68-0.83~(0.72\pm0.01)$ mm. Width at the base of proboscis was $0.10\pm0.02~(0.08-0.12)$ mm. Proboscis club-shaped with a broad anterior end, equipped by longitudinal rows of hooks, each with 15-19 of curved hooks. Trunk was spines anteriorly; spines arranged in 9-11 collar rows, each was equipped with 15-18 spines. Whited showed that the parasite isolated is belonged to be *Serrasentis sagittife*. These postmortem findings are similar to that recorded by **Abdel-Mawla and AboEsa (2011) and Salah Eldeen** *et al.* **(2014).**

Serrasentis sagittifer

Description (Based on 7 Specimens): helminthe were creamy white, elongated, and curved with slight tegument annulations and a narrow posterior end. It was characterized by the presence of distinctive rows of spines (combs) on the ventral surface of the trunk. And measured 5.8-9.7 $(6.9\pm0.2)\times0.52-0.69$ (0.58 ± 0.02) mm for male and 10.1-11.3 $(10)\times0.68-0.83$ (0.72 ± 0.01) mm for female. Proboscis was long and cylindrical with a length of 0.89-1.3 mm (1.1 ± 0.2) for male and 1.11-1.19 mm (1.17 ± 0.02) or female It was covered with numerous uniform spines arranged longitudinally as 9 to 11 rows each equipped by 15 to 18 spines. **Table 1**.

Taxonomic Summary Parasitological finding:

Phylum: Acanthocephala

Family: Rhadinorhynchidae Travassos (1923)

Subfamily: Serrasentinae

Species: Serrasentis sagittifer Fig. 1, 2

Type Host: Red porgy *Mullus barbatus* (<u>Mullidae</u>)

Infection Site: pyloric ceca, Intestine, body cavity, mesenteries and external surfaces of

internal organs.

Locality: EL-koms coasts, the Mediterranean Sea, Libya.

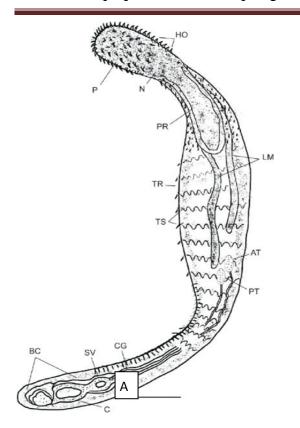




Fig.1 A: *Serrasentis sagittifer* (male), lateral view, P, proboscis; pr, proboscis receptacle; N, neck; HO, hooks; L, lemnisci; TS, trunk spines; T, trunk; AT, anterior testis, BC, bursa copulatrix; PT, posterior testis; CG, cement gland, SV, seminal vesicle, C, cirrus; bar, 100μm. **B**: showing scanning Electron: Proboscis and multiple rows of Proboscis hooks at, 25 kV.





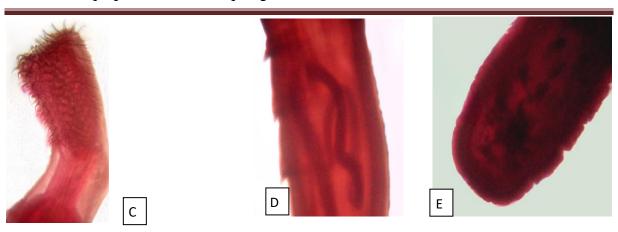


Figure 2. Photomicrographs of Serrasentis sagittifer larva.

A and B: Whole mount of juvenile

C: The bodyD: trunk spinesE: Posterior end

Table 1: Comparative data (mm) of males and Females *Serrasentis sagittifer* recorded in the present study and those of other previous studies.

pecies	Serrasentis	Serrasentis	Serrasentis	Serrasentis
pecies	sagittifer	sagittifer	sagittifer	sagittifer
Referance	AL- Zubaidy and	Abdel-Ghaffar et	Saed et al, (2018)	Present study
Refer ance	Mhaisen(2012)	al.(2014)	Saca et al, (2010)	1 Tesent study
Host	Thunnus tonggol	Sparus aurata	Saurida	Mullus
HUSt	Sphyraena	sparus auraia	undosquamis	barbatus
	barracuda		инаоздиатиз	barbaius
	Pomadasys			
	argenteus and			
	Lutjanus gibbus			
Body Length	7.5-9.0	6.9-8.6	3.33-3.58	males
V 6				5.8–9.7 mm
Body width	0.62-0.75	0.57-0.73	0.08-0.12	0.52–0.69 mm
Proboscis length	1.0-1.3	0.97-1.6	0.71-0.77	0.89–1.3 mm
Number of rows	22	9-11	15-19	9–11
Number of	16	15-18	15-18	15–18
spines				
				Females
Body Length	-			10.1–11.3 mm
Body width	-			0.68–0.83 mm
Proboscis length	-			1.11–1.19 mm
Number of rows	-			9–11
Number of	-			15–18
spines				
Locarion	Red Sea, Yemeni	Coasts of the Red	Coasts of the Red	Coasts of the
	coastal waters	Sea at Hurghada	Sea at Hurghada	Mediterranean
		City, Egypt	City, Egypt	Sea at
				AL.kuoms
				City, Libya

Discussion

in the current study, Serrasentis sagttifer were collected from the intestine, pyloric caeca and peritoneal cavity of *Mullus barbatus*. The total prevalence of Serrasentis sagittifer in the Mullus barbatus fishes were 24.16%. Meanwhile, the obtained result was different from that reported by (Abdou and Mahfouz 2006) which was 33% acanthocephala from Siganus luridus (Salah Eldeen et al., 2014), 38% clerocollum sp from Siganus revulatus. The spread of Acanthocephala (Echinorhynchus and Serrasentis sagttifer) from Mulloides sp flavolineatus was 23.3% which was nearly similar to the results reported by (El-Ashram and Shager 2008), 25% Serrasentis sagttifer from Scombermorus maculates and (Al Zubaidy and Mhaisen 2012) 24% from Pomadasys argenteus fish. (Abo-Esa 2007) wsa 35% S. sagttifer from Mullus barbatus. Moreover, (El-Ashram and Shager 2008) showed 25% S. sagttifer from Scombermorus maculates and lower than that investigated by Abdel-Ghaffar et al., (2014) which showed a higher percentage (57.14%) of S. sagittifer from Sparus aurata at the coasts of the Red Sea and lower than that investigated by (Kassem et. al., 2023) showed a higher percentage (48.5%) and (Faisal et. al., 2020) showed the total prevalence of Serrasentis sagittifer (100%). This variation in prevalence may be Attributed to the unequal samples, difference of fish species, species of parasites and different sites from which samples collected as well.

In the meantime, a higher percentage than what was obtained by Al-Zubaidy and Mhaisen (2012) who reported an overall prevalence of 13.1% in some marine fishes and Bayoumy et al, (2008) 16.2% *Echinorhynchus gadi* from *Mullus surmuletus*, (Abdel-Mawla and El-Ekiaby 2012) 7% from Seabass fish and Debenedetti et al, (2013) 3% parasite *Echynorhynchus gadi* from *Mullus barbatus*, While recording Öztürk and Yeşil (2017) 1.21% parasites Acanthocephaloides from Red Mullet.

This morphological description agrees with that described by Al-Zubaidy and Mhaisen (2012), Barton and Smales 2015 and Abdel-Mawla and El-Lamie 2018).

To conclude, the *S. sagittifer* in the present study, morphologically resembles those previously recorded from various regions of the world **AL-zubaidy & mhaisen**,

(2012); Abdel Ghaffar *et al.*, (2014); Mohamadain & Adel (2015); Çelik and Oguz, 2021 and Kassem *et al*, (2023).

References

Abdou, N.E., & Mahfouz, M.E. (2006): Ultrastructural and genetic diversity studies of two Sclerocollum (Acanthocephala) sp. Infecting siganid and lutianid fishes from Red Sea, *Egypt. J.Egypt. Soc. Parasitol.* Dec; 36 (3), 1035-56.

Abo-Esa, F. K. Jihan (2007). Helminth parasites in Barbony *Mullus barbatus* fish with reference to public health hazards. *Egypt. J. Aquat. Biol. & Fish.*, 11(3): 127-137.

Abdel-Mawla, I. Heba & Abo-Esa, F. Jihan (2011): The most common parasitic diseases in *Siganus revulatus* in Suez Canal area. *The Egyption Vet. Med. Assoc.*, 71(1): 257-270.

Abdel-Mawla, I. Heba & El-Ekiaby, T. Walaa, (2012). Some studies on parasitic infection among *Morone labrax* (Seabass fish) as bio-indicator of environmental conditions. *Egyptian Journal for Aquaculture*, 2 (3):1-15.

Abdel-Mawla, I. Heba & El-lamie, M. Maather (2018). Investigation of Acanthocephalan parasites in some marine fishes as a bio-indicator for heavy metals pollution. *Egyptian Journal for Aquaculture*, 8 (1):13-30.

- **Abo-Esa, F. K. Jihan (2007).** Helminth parasites in *Barbony Mullus* barbatus fish with reference to public health hazards. *Egypt. J. Aquat. Biol. and Fish*, 11(3): 127-137.
- **Al-Zubaidy, A.B. & Mhaisen, F.T.** (2012). A record of two species of Acanthocephala (Echinorhynchida: Rhadinorhynchidae) from Red Sea fishes, Yemeni coastal waters. *Mesopot. J. Mar. Sci.*, 27 (1): 15 28.
- **Abdel-Ghaffar, F.; Morsy, K.; Abdel-Gaber, R.; Mehlhorn, H.; Al Quraishy,S. & S. Mohammed (2014)**. Prevalence, morphology, and molecular analysis of *Serrasentis sagittifer* (Acanthocephala: Palaeacanthocephala: Rhadinorhynchidae), a parasite of the gilthead Sea bream *Sparus aurata* (Sparidae). *Parasitol. Res.*, 113:2445–2454.
- **Bush, A.O., K.D. Lafferty, J.M. Lotz & W. Shostak** (1997). Parasitology meets ecology on its own terms: Margolis et al. revisited. *Journal of Parasitology*, 83: 575-583.
- **Barton D. P. & Smales L. R. (2015).** Acanthocephalan cystacanths from flatfish (order: pleuronectiformes) in tropical Australian waters. *J. Parasitol.*, 101(4), pp.429 435.
- **Bayoumy, M. E.; Abd El-Monem, S. & Ammar, A. K. (2008).** Ultrastructural study of some helminth parasites infecting the Goatfish, *Mullus surmuletus* (Osteichthyes: Mulldae) from Syrt coast, Libya. *Nature and Science*, 6(2): 51-63.
- **Chambers, C.A. & Dick, T.A.** (2005). Trophic structure of one deep-sea benthic fish community in the eastern Canadian Arctic: Application of food, parasites and multivariate analysis. *Environmental Biology of Fishes*, 74(3-4): 365-378.
- ÇELİK B. A. & OĞUZ M. C. (2021). Endohelminth fauna of teleost fishes from coasts of Şile region of the Black Sea. *HELMINTHOLOGIA*, 58, 3: 263 270.
- **Debenedetti, A.L.; Madrid, E. and Fuentes, M.V.** (2013). Study of helminth parasites in the red mullet, Mullus *barbatus*, from the Mediterranean Sea and acquired in greater València, Spain. Rev. *Ibero-Latinoam. Parasitol*; 72 (2): 118-123.
- **Dogiel, V.A.** (1964). *jeneral parasitology*. London: Oliver and Boyd.
- **El-Ashram, A. M. M. & Shager, G. E. (2008).** Studies on enteric parasitic diseases caused by prevailing helminthes among some marine fishes from the Red Sea. Abbassa Int. *J. Aqua*; 16: 415 444.
- Faisal D, Yousef N. S. I. Y, Abdel-Mawla H.I. & El-Kalamawy N. (2020). Acanthocephalan Infection in Rachycentron Canadum Fish in Red Sea. j. Egypt. vet. med. Assoc 80, no 1, 39-53
- **Gibson, J.W.,** (1985). Satisfaction with Upward and Downward Organizational Communications: Another Perspective. Proceedings of the Southwest Academy of Management (March): 150.
- **Kassem H., EL-Maadani R, Bojwari G., Bowashi S. & Hador A. (2023).** Nematode infestation in Red Mullet (*Mullus surmuletus*, Linnaeus, 1758) From Benghazi Coast, Libya. Al Qalam *Journal of Medical and Applied Sciences*; Volume 6, Issue 1.
- **Mohamadain H.S., & Adel A**. (2015). Light and scanning electron microscopy on *Serrasentis sagittifer* Linton, 1889 (Acanthocephala): Palaeacanthocephala: Rhadinorhynchidae) infecting the common sea bream in Egypt. *J Egypt Soc Parasitol* 2015; 45(1): 23-28. http://dx.doi.org/10.12816/0010846. PMid: 26012215.
- Öztürk T. & Yeşil A. (2017). Metazoan parasite fauna of the Red Mullet, *Mullus barbatus* ponticus Essipov, 1927 in the Sinop Coasts of the Black Sea. Turkish *Journal of Fisheries* and Aquatic Sciences 18: 153-160.
- Sanil, N.K.; Asokan, P.K.; Lijo John & Vijayan, K.K. (2010). Pathological manifestations of the acanthocephalan parasite, Tenuiproboscis sp. In the mangrove red snapper (*Lutjanus*

argentimaculatus) (Forsskål, 1775), a candidate species for aquaculture from Southern India. *Aquaculture*, 310:(259-266), Issues 3-4.

Sakthivel, A.; Selvakumar, P. & Gopalakrishnan A. (2014). Acanthocephalan (*Echinorhynchus* sp.) infection of yellowfin tuna (*Thunnus albacares*) from Nagapattinam, south east coast of India. *Journal of Coastal Life Medicine*; 2(8): 596-600.

Salah Eldeen, Y.M.H.; Idris, O. F.; Sabahelkhier, M.K. & Abdelhaleem, M. I. (2014): Histopathological alterations in small intestine of rabbit fish (*Siganus rivulatus*) infected by helminth parasite (*Sclerocollum* sp.), red sea coast, sudan. International *Journal of Environment*, 3, (216-228), Issue 2.

Saed, N., Mahrashan Abdel-Gawad, Sahar El-Ganainy, Manal Ahmed, Kareem Morsy & Asmaa Adel (2018). *Serrasentis Sagittifer* Linton, 1889 (Acanthocephala: Rhadinorhynchidae) from the Red Porgy *Pagrus pagrus* (Teleostei: Sparidae) of the Red Sea, Egypt: A Morphological Study Acta Parasitologica Globalis 9 (3): 133-140.

Nickol, B.B. (2006). Phylum Acanthocephala, In: Woo, P.T.K. (Ed.), Fish Diseases and Disorders, Second Edition.: Protozoan and Metazoan Infections, I (444–465). CAB International, Wallingford, UK.

Taraschewski, H. (2000). Hostparasite interactions in Acanthocephala: A morphological approach. *Advances in Parasitology*, 46:1-179.

Tingbao, Y. & Xianghua, L. (2001): Seasonal population dynamics of *Neoechinorhynchus qinghaiensis* in the carp, *Gymnocypris przewalskii* przewalskii, from Qinghai Lake China. *Journal of Helminthology*, 75(1): 93-98.

i